

# Antibiotics from Marine Bacteria

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**Abstract**—This review discusses main directions and results of the studies on antibiotics produced by bacteria living in the marine environment. In recent years many obligate marine species and strains were studied, diverse metabolites were isolated, and their chemical structures were elucidated. Among them here were natural compounds toxic against tumor cells, pathogenic bacteria, viruses, and malaria plasmodial species; these compounds often had no analogues among the natural products of terrestrial origin. Some isolated compounds form a basis of active ingredients in medicinal preparations used in clinic practice, while others are under different stages of preclinical or clinical studies. Much attention has been paid in recent years to producers of marine-derived antibiotics isolated from the deep-sea habitats, from the surface of marine invertebrates and algae, as well as from symbiotic microorganisms.

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## INTRODUCTION

The discovery and widespread use of antibiotics had been of paramount importance for the fate of mankind and significantly reduced the number of deaths of epidemics that claimed lives of millions of people for many centuries. It is believed that this was one of the main causes for the increase in human life expectancy. For example, in Russia at the end of the 19th century, it was about 30 years, by 1961-1962 it had grown to 67.85 years, and now life expectancy is about 73.4 years (2018). However, the main natural source of antibiotics, soil microorganisms, has now significantly exhausted its potential, and the number of new antibiotics isolated from soil bacteria is decreasing.

At the same time, the search for new antibiotics remains an important task due to the increasing number of cases of severe infections caused by various pathogens, primarily drug-resistant strains. As a result, the number of patients with drug-resistant diseases is increasing. In 2017, 10 million people were diagnosed with tuberculosis, and 1.6 million people (including 0.3 million people with HIV) died from this disease [1]. In 2008 a group of pathogens associated with the severe drug-resistant infec-

tions was identified, which was named “ESCAPE BUGS” (acronym of the Latin names of the following antibiotic insensitive pathogens: *Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Enterobacter* spp.) referring to their ability to ESCAPE the effects of the commonly used antibiotics. Later, properties of these pathogens were described in more detail [2]. The threat of infection with antibiotic-resistant strains of several other pathogenic bacteria, including *Helicobacter pylori*, *Campylobacter* spp., *Neisseria gonorrhoeae*, and others also increased in recent years [3].

The urgent need to find new antibiotics forced scientists to pay attention to the natural sources of these biologically active substances that are difficult to study, especially such as marine microorganisms – bacteria and microscopic fungi – that live under extreme conditions, namely to the so-called extremophiles (barophiles, psychrophiles, thermophiles, etc.), as well as epiphytic and symbiotic microbial flora of invertebrates and algae. Using new opportunities to search for antibiotics, including descent underwater vehicles and specialized ships, scientists have found antibacterial, antifungal substances and compounds that inhibit pathogenic protozoa in such non-traditional producers of antibiotics as animals, especially marine invertebrates, inhabitants of underwater hot

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springs, searched for these substances in the Arctic and Antarctica, at the maximum depths of the ocean, in deep caves and deserts. As a result, a series of substances with anti-pathogenic properties was identified, which can be considered as leader compounds for design of new drugs.

To date, several review articles have been published on various aspects of the search for antibiotics isolated from marine microorganisms (see [4-10] for example) describing more than a hundred of such compounds. The reviews concerning isolated metabolites that inhibit methicillin-resistant *S. aureus* [11] and vancomycin-resistant enterococci were also published [11, 12].

Although a significant number of marine microorganisms have not yet been cultured, other marine microorganisms have been studied and appeared to be quite different in their physiological and morphological features when compared with their terrestrial counterparts. These peculiarities help them to adapt and survive in the seas and oceans. Among marine microorganisms, there are many previously unknown species, genera, and even families of bacteria, and their discovery has greatly changed modern microbiology. For instance, several hundred new species of bacteria were validly described by scientists from our Institute under the leadership of Professor V. V. Mikhailov – corresponding member of the Russian Academy of Sciences. Like other marine bacteria, they produce a wide variety of bioactive metabolites [10, 13]. The first accurately described new species of heterotrophic bacteria in the Russian Far East region was *Alteromonas fuliginea* (type strain KMM 216T), inhabiting waters of the Peter the Great Bay (1995). Later, this species was assigned to the newly established genus *Pseudoalteromonas* and included in the species *Pseudoalteromonas citrea*. Over the next ten years, *Marinomonas primoryensis* sp. nov. (named in honor of the Primorye Region, Russia), *Vitellibacter vladivostokensis* gen. nov., sp. nov. (named in honor of Vladivostok), *Arenibacter troitsensis* sp. nov. (named in honor of Trinity Bay, in which waters this species lives, and on the shores of which the Marine experimental station (MES) of the PIBOC is located), *Mesonina algae* gen. nov., sp. nov. (named in honor of MES), *Salinibacterium amurskyense* gen. nov., sp. nov. (these bacteria were isolated from a sample of seawater from the Amursky Bay), *Reinekea marinisedimentorum* gen. nov., sp. nov. (named in honor of the Reinecke Island near Vladivostok, in waters of which the corresponding microorganism was found in the sandy soil at a shallow depth) and others were described. Only during the period from 1995 to 2005, more than 100 new species, about 40 new genera, and one new family of marine bacteria were discovered by the scientists working at the Microbiology laboratory of PIBOC [13]. Some of them produce a variety of bioactive compounds, including antibiotics [14]. To date, the number of new species and genera in the collection of marine microorganisms (acronym is KMM) of the PIBOC has increased significantly.

However, not so long ago, in 1996, only 7 genera of bacteria were considered exclusively marine, such as *Oceanospirillum*, *Marinomonas*, *Planococcus*, *Listonella*, *Leucothrix*, *Platobacterium*, and *Prochloron* [4]. Many of them are associated with marine fish, invertebrates, or algae. Later, the situation changed and the number of such taxa increased. Moreover, taxa of a higher taxonomic rank were found, all the representatives of which were obligate marine organisms. On the other hand, even in the cases when bacteria isolated from the sea water or sediments were found to belong to species previously isolated from the terrestrial biological sources, they could not be considered simply as washed away from the land and preserved all their properties in the new environment. Adaptation to the marine conditions often led to the selection of such metabolic pathways that generated previously unknown metabolites. This explains the significant biochemical diversity in these microorganisms and contributes to their study as new sources of antibiotics.

Actinobacteria are the most known bacterial producers of antibiotics. They belong to the family Actinomycetaceae, which includes the genera *Streptomyces*, *Actinobaculum*, *Acanabacterium*, and others. Most of these Gram-positive bacteria produce compounds with unique chemical structures that are toxic against other bacteria, including pathogenic ones. More than 60% of all antibiotics produced by the pharmaceutical industry were obtained by culturing actinobacteria [15, 16]. These include streptomycin, actinomycin, polyenes, tetracyclines, macrolides, and other medications. It is believed that each strain of actinobacteria, due to its genetic potential, is capable of synthesizing 15-25 different secondary metabolites. It was estimated that about 10 thousand new bioactive substances of various chemical natures could be found in actinobacteria [16].

For many decades, terrestrial sources, mainly soil microorganisms, have been used as a source for searching and designing new antibiotics. Although as early as in 1969, there was a report about the first mycelium-forming actinobacteria from marine bottom sediments [17]. Moreover, it has been suggested that marine antibiotics, including those isolated from symbiotic microorganisms of marine invertebrates, will have an advantage in creating new drugs [17, 18].

Among the various genera of actinobacteria, the genus *Streptomyces* is of particular interest, because it is known as a source of more than half of all antibiotics currently used in clinical practice. 5200 genomes of these actinomycetes are registered in the Genome Online Database [19]. Interestingly, new species of this genus have been discovered in recent years, which can be considered exclusively marine streptomycetes [20]. Moreover, 5 new genera of marine actinomycetes have been recently described: *Marinactinospora* [21], *Jonesia* [22], *Salinibacterium* [23], *Sciscionella* [24], and

*Salinispora* [25]. Many new bacteria, belonging to other taxonomic groups were also discovered, and highly active secondary metabolites were found in them.

In this short review, we will provide information about the main directions of the search for marine-derived antibiotics and illustrate these directions only with a small number of examples of promising marine-derived antibiotics, grouping them by their inhibitory effects against tumor cells, pathogenic bacteria, fungi, and other pathogens.

### CYTOTOXIC ANTIBIOTICS FROM BACTERIA ISOLATED FROM MARINE BIOLOGICAL SOURCES

The search for marine antibiotics that exhibit an inhibitory effect against tumor cells has been actively carried out in several countries in recent years, especially in the United States and Japan. *Salinispora* bacteria have attracted much attention from microbiologists, bioorganic chemists, and pharmacologists. Their cultivation was first performed in 1989 after isolation from the samples of bottom sediments collected at a depth of 1100 meters. These bacteria, which originally were assigned to the genus *Micromonospora*, did not grow if sea water was replaced with ordinary deionized water in the medium. As *Salinispora*, they were described in 2005 [25]. After the cultivation technique was improved, *Salinispora* spp. were also found in some marine invertebrates (sponges, ascidia) and in algae. Bacteria of this genus turned out to be obligately marine species, and their further study led to the discovery of several antibiotics that exhibit strong antitumor effect [26].

The first group of interesting metabolites from this species were obtained from the bacteria isolated from a pre-heated sample of bottom sediments. The main obtained substance was called salinosporamide A (**1**) [27]. This new antibiotic has a skeletal system consisting of  $\gamma$ -lactam and  $\beta$ -lactone (Fig. 1) and demonstrates cytotoxic properties against tumor cells. Although its cyclic system is identical to that of omuralide (**2**), isolated by Japanese microbiologists in 1991 from *Streptomyces lactacystennaosus*, the compound differs significantly from the omuralide (**2**) in its substituents and their positions. Salinosporamide A proved to be a powerful and selective inhibitor of proteasomes; it is approximately 35-fold more active in this test than omuralide. The antibiotic inhibits cell lines of lung cancer NCI-H226, brain tumors SF-539, melanoma SK-MEL-28 and MDA-MB-43, with  $IC_{50}$  values (concentrations that inhibit cells by 50%) in these tests being less than 10 nM. Salinosporamide proved to be an efficient anti-cancer agent in preclinical trials, and is undergoing human clinical trials as a preparation against melanoma. The corresponding drug called Marizomib is under development

by “Nereus Pharmaceutical” (San Diego, USA) under a license obtained from the University of California, San Diego.

Different strains of *Salinispora* spp. produce also other antibiotics. For instance, the study of the *S. tropica*, strain CNB-392, led to isolation of salinosporamides B (**3**) and C (**4**), while replacement of NaCl with NaBr in the medium produced the compound (**5**) (Fig. 1), which has a bromine atom instead of chlorine in the side chain and retains cytotoxic properties [28, 29].

Cyclic peptides from the bacterium *Streptomyces* sp. (strain CNQ-593) from marine bottom sediments collected at a depth of 20 m near the Guam Island are also considered promising for cancer treatment. These substances were called piperazimycins and have formulae (**6-8**) (Fig. 1) [30]. Piperazimycins show strong toxic properties against tumor cells. The average cell growth-inhibitory concentration ( $GC_{50}$ ) after testing on 60 tumor cell lines, provided by the National Cancer Institute, USA, was approximately 100 nM.

In many cases, highly active metabolites from marine bacteria exhibit both cytotoxic and antibacterial properties, although sensitivity of the tumor cells and pathogenic bacteria to them could be different. For example, new derivatives of aureolic acid, chromomycins A<sub>2</sub>-A<sub>4</sub> (**9-11**) (Fig. 1), obtained from *Streptomyces* sp. (strain KKM 9048, which was isolated from a sediment sample collected in the North-western part of the Sea of Japan), showed strong antibacterial action against *Enterococcus faecium*, *Staphylococcus aureus*, *S. epidermis*, and *Bacillus subtilis*. However, their inhibitory concentrations against tumor cells were even lower, especially those that inhibit formation of microcolonies of RPMI-7951 and SK-Mel-28 tumor cells (10-20 nM) [31].

The marine-derived isoquinoline derivative, ecteinascidin-743 (ET-743) (**12**) (Fig. 1), became the active substance of the new anticancer drug Trabectedin (Yondelis R), developed by the Spanish company “PharmaMar” (manufacturer “Baxter Oncology GmbH”, Germany). This substance itself was discovered in the USA at the University of Illinois by Prof. K. Rinehart. He was able to isolate it from the ascidian *Ecteinascidia turbinata* and demonstrate its very high antitumor activity [32]. The Nobel Prize laureate E. J. Corey and collaborators synthesized compound **12**, but later a semisynthetic process was developed by PharmaMar, starting from the antibiotic safracin B. Since 2007, Trabectedin has been used in clinical practice for treatment of sarcomas, ovarian cancer and other malignant tumors.

Recently, it was found that this ascidian contains a consortium of various symbiotic bacteria with the main species being the  $\gamma$ -proteobacterium *Candidatus Endoecteinascidia frumentensis*. The use of Meta-Omics technology allowed characterizing this consortium and

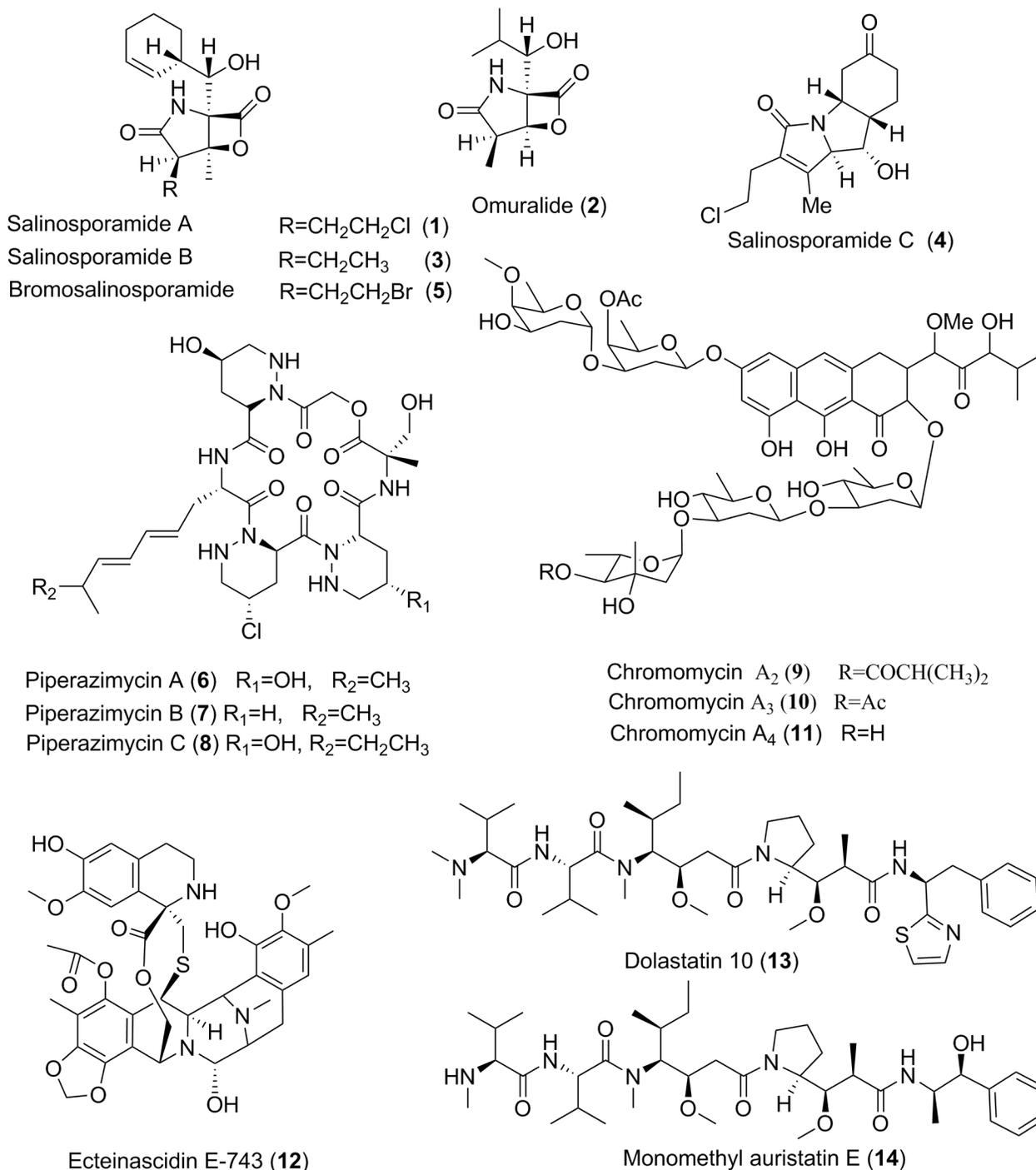


Fig. 1. Some cytotoxic metabolites of marine bacteria.

identifying 25 genes that catalyze formation of ET-743. Subsequent sequencing showed that the true producer of the active substance of Trabectedin is this symbiotic bacterium [33].

Actinobacteria are the most prevalent among the invertebrate symbiont bacteria. They appear to be involved in biosyntheses of many metabolites from

marine invertebrates, including those that have been used as active substances in medicinal drugs [34]. Other symbiotic bacteria have also been found in marine invertebrates. For example, another large group of symbiotic bacteria, is present in sponges that participates in biosynthesis of various secondary metabolites. This group belongs to a higher-rank taxon – representative of a novel

candidate phylum Poribacteria [35]. In addition, symbionts of marine invertebrate have been also identified as cyanobacteria, firmicutes, and microscopic fungi.

Cyanobacteria are known for their extremely active peptides, including dolastatins, the most potent from all the natural and synthetic compounds known to be toxic against tumor cells. They inhibit growth of tumor cells in picomolar concentrations. Dolastatins were first isolated from the mollusk *Dolabella auricularia*, but their genuine producers proved to be the cyanobacteria *Symploca hydroides* and *Lyngbya majuscula*. A derivative of dolastatin 10 (**13**), the so-called monomethyl auristatin E (**14**) (Fig. 1), was selected as an active substance for designing of the anticancer drug Vedotin (Brentuximab or Adcetris<sup>R</sup>) – representative of a new generation of targeted anticancer drugs. This is a conjugate of monoclonal antibodies with above-mentioned derivative of powerful marine-derived cytotoxin. Vedotin has been used since 2011 for treatment of lymphomas and other oncological diseases [36, 37], manufacturer – “BSP Pharmaceutical S.r.l.”, Italy.

#### MARINE ANTIBIOTICS WITH ANTIBACTERIAL PROPERTIES

However, majority of the marine-derived antibiotics demonstrate strong antibacterial effect. The search for and discovery of these natural products are of great importance, since infectious diseases remain one of the main causes of death. Among them, there are quite different compounds in chemical structures: heterocyclic derivatives, quinoids, products of polyketide origin, peptides, etc. Dozens of research teams from many countries are actively searching for such substances. The laboratory of Professor William Fenical from the Scripps Institute of Oceanography (San Diego, USA), discovered several other marine antibiotics in addition to salinosporamides, for example anthracimycin (**15**) with a strong inhibitory effect against pathogenic bacteria. This antibiotic, isolated from the actinobacterium *Streptomyces* sp., strain T676, collected off the St. Johns Island near Singapore, inhibits *Bacillus anthracis*, the causative agent of anthrax, which causes a deadly disease and has already been used by terrorists who sent letters with spores of this bacterium to their targets. However, for the first time anthracimycin was obtained from another marine streptomycete, strain CNH365, collected from the bottom sediments off the coast of Santa Barbara, California, USA [38]. Chemical structure of the compound **15** was established by careful analysis of its NMR spectra and confirmed by X-ray diffraction analysis. Interestingly, its structure has an unusual hydrogen bond formed by two ketone groups with one of them in enol form (Fig. 2). When treated with N-bromosuccinimide in dichloroethane, anthracimycin forms a dichloro derivative (**16**), which also shows high antibac-

terial activity, especially against methicillin-resistant *E. coli* and some other pathogenic bacteria.

The anthracimycin biosynthesis was studied at the genetic level. The identified “atc” gene cluster responsible for its biosynthesis consists of 53253 base pairs. It includes the genes of type I polyketide synthase and acyltransferase, which catalyze formation of malonyl and methylmalonyl coenzymes A involved in the construction of the skeletal system of **15**. Expression of this gene cluster in the bacterium *Streptomyces coelicolor* led to the production of anthracimycin in this bacterium. 10 biosynthetic modules participate in this biosynthesis, the decalene cyclic system is first formed under their action, and after it the biosynthesis is completed by a thioesterase that catalyzes closure of the macrocycle to form a lactone bond [39].

New antibacterial agents (**17**, **18**) (Fig. 2) were found in the *Streptomyces* sp. marine isolate from cyanobacterial mat collected off the coast of Puerto Rico. These unusual bis-anthraquinones show strong inhibitory effects against strains of vancomycin-resistant *Enterococcus faecium*, methicillin- and tetracycline-resistant *Staphylococcus aureus*. In addition, their synthetic derivatives have been also studied. The most effective antibiotic (**17**) demonstrates 50% bactericidal concentrations (MIC<sub>50</sub>) of 0.11, 0.23, and 0.90 µg per ml against the above-mentioned dangerous bacterial pathogens [40].

The bacterial strain NPS12745 was isolated from the sediment samples collected from the bottom of Missoni Bay in San Diego, California. Based on the nucleotide sequence of its 16S RNA gene, this bacterium was identified as a new species belonging to the genus *Marinispora*. The extract of this microorganism inhibits growth of both Gram-positive and Gram-negative pathogenic bacteria. The structures of the obtained antibiotics were established as new bis-indolyl pyrrole derivatives called lynamicins A-E (**19-22**) (Fig. 2). All of them demonstrated strong antibacterial effect. Lynamicin B showed the highest activity, and it inhibited strains of pathogenic bacteria *Streptococcus pneumoniae*, *Haemophilus influenzae*, and *Staphylococcus aureus* at concentrations of 2–4 µg/ml [41].

Gageomacrolactins such as gageomacrolactin (**23**) (Fig. 2) from the bacteria isolated from the bottom sediments on the Gageo reef in the waters of the Republic of Korea, showed high antibacterial activity against both Gram-positive (*Staphylococcus aureus*, *Bacillus subtilis*) and Gram-negative bacteria (*E. coli*, *Salmonella typhi*, *Pseudomonas aeruginosa*) [42].

Peptide antibiotics were quite often found in the different marine bacteria, such as actinobacteria and cyanobacteria. Screening of 44 marine actinomycetes from the family Micrococaceae, isolated from sponges collected off the coast of Florida (USA), led to the discovery of new producers of antibiotics. They were identified by genetic methods as representatives of the genera

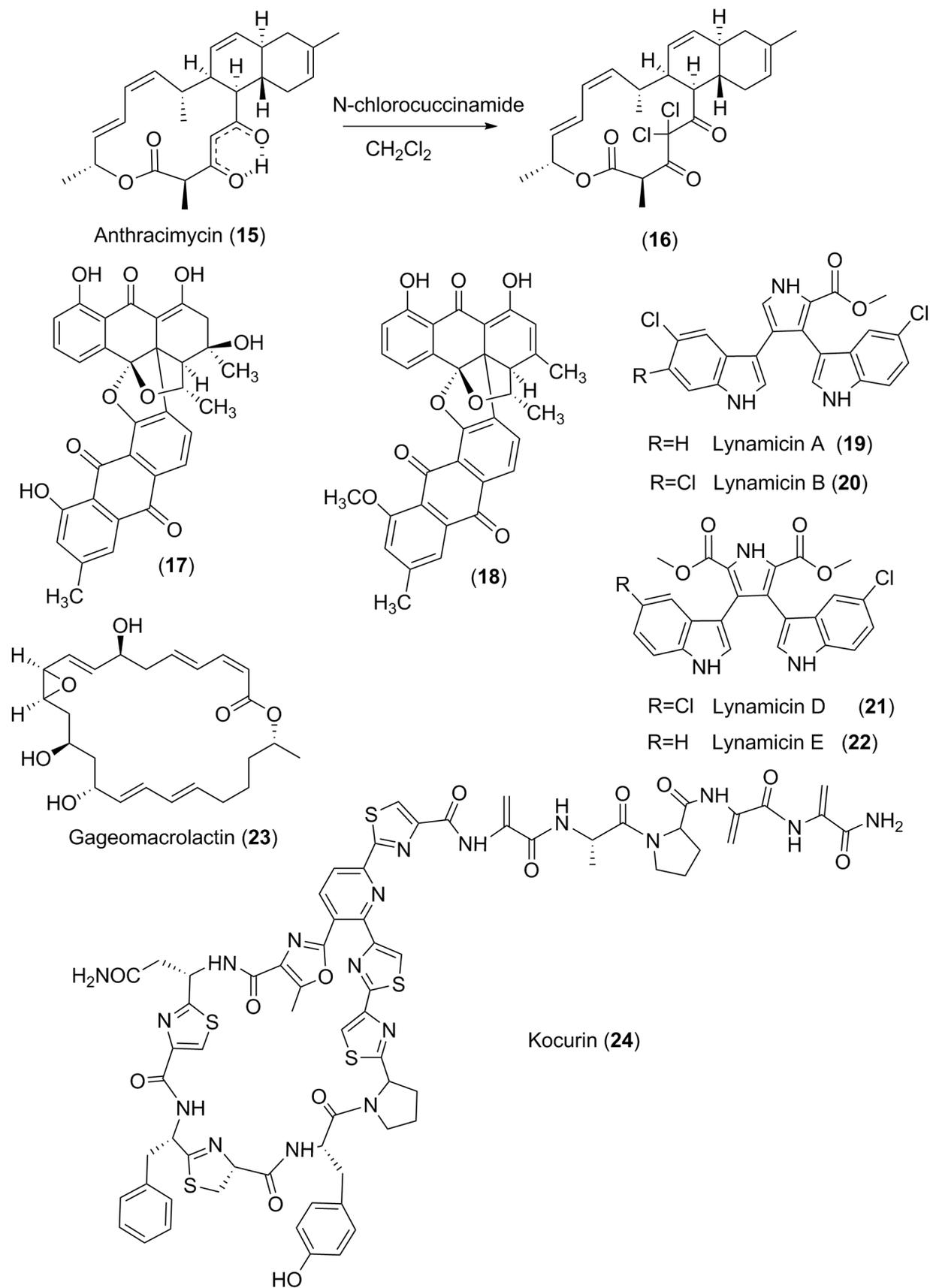


Fig. 2. Some antibacterial metabolites of marine bacteria.

*Kocuria* and *Micrococcus* and contained the genes of non-ribosomal peptide synthase and polyketide synthase. Three obtained isolates produced the peptide antibiotic Kokurin (**21**) (Fig. 2) – a new member of the family of thiazole-containing cyclic peptides [43].

A compound with similar structure, called antibiotic PM181104 (Fig. 2), was isolated by the Indian chemists after cultivation of a bacterium belonging to the same genus isolated from the sponge *Spirastrella inconstans* var. *digitate*, collected in Palk Bay, India. This antibiotic inhibits various pathogenic bacteria, including antibiotic-resistant strains, at concentrations of approximately 1 nM. In *in vivo* experiments it provided 100% protection of animals against sepsis caused by the methicillin-resistant *S. aureus* and vancomycin-resistant enterococci when administered into mice at doses of 5 or 10 mg/kg, respectively [44].

The number of promising antibiotics from marine organisms is increasing year after year, which attracts attention of more and more research groups around the world.

#### ANTIFUNGAL MARINE ANTIBIOTICS

Pathogenic fungi that cause severe mycoses remain one of the main threats to the cancer patients with weakened immune systems as a result of chemotherapy, as well as to the patients with AIDS and other diseases that initiate immunodeficiency. Such patients are often infected with opportunistic fungi such as representatives of the genera *Candida*, *Aspergillus*, and *Cryptococcus* leading to serious consequences up to fatal outcomes. About 500 thousand people die from such infections every year, and economic impact is about 12 billion dollars. The search for natural antifungal compounds and development of drugs on their basis is an urgent task of modern science.

Some metabolites of marine bacteria, particularly actinomycetes, have become known for their antifungal properties in recent years. For example, saadamycin (**25**) (Fig. 3), a polyketide antibiotic from the actinobacterium *Streptomyces* sp. (a new species of this genus), isolated from the sponge *Aplysina fistularis* collected off the coast of Egypt, has a powerful antifungal effect against *Candida albicans*, *Aspergillus* spp. and *Cryptococcus* spp. with a minimum inhibitory concentrations (MIC) of 1–5.2 µg/ml and a minimum fungicidal concentrations (MFC) of 2–10 µg/ml [45]. Interestingly, scientists who discovered this new microbial source were able to increase production of the antibiotic 2.26-fold by treating its producer with ultraviolet rays. The resulting mutant strain Ah22 was then used to isolate and establish the structure of antibiotic itself. The structure of the compound **25** is not very complex. It is a product of polyketide biosynthesis that has an  $\alpha$ -pyrone cycle, and proved to be a derivative of 5-hydroxy-2-oxo-2H-pyrane.

A phenoxazine derivative, chandrananimycin A (**26**) (Fig. 3), and related compounds were isolated from the actinobacterium *Actinomadura* sp. collected from the bottom sediments of the Bay of Guangzhou (China). Compound **26** inhibits a number of fungi, including *Mucor miehei*, whose enzymes are used in cheese production [46].

A series of actinomycetes, including the BM-17 strain, were isolated from a sample of bottom sediments collected in the Arctic ocean. This strain has been identified as *Nocardia dassonvillei*. A new secondary metabolite (**27**) (Fig. 3) along with previously known phenazine antibiotics, was obtained when this strain was cultured. This compound shows significant antifungal activity against *Candida albicans* with MIC<sub>50</sub> of 64 µg/ml and potent cytotoxic properties against a number of tumor cell lines [47].

Other marine macrolides also often show antifungal properties. For example, macrolactin A (**28**) (Fig. 3) and similar macrolactins isolated from the bacterium *Bacillus subtilis* collected in the area of the above-mentioned Gageo reef exhibit strong antifungal action against *Aspergillus niger*, *Candida albicans*, *Botrytis cinerea*, *Colletotrichum acutum*. Minimum inhibitory concentrations were as low as 0.04–0.3 µM [48]. Some related macrolides also exhibit potent antifungal effects [49].

Neomaclafungin A (**29**) (Fig. 3) from the bacterium *Actinoalloteichus* sp. NPS702, isolated from the sample of bottom sediment collected in Usa Bay (Japan) [50], shows significant antifungal activity against *Trichophyton mentagrophytes* (MIC 1–3 µg/ml). After synthesis, its structure was revised [51].

Like bacteria, pathogenic fungi also develop drug resistance mechanisms and form fluconazole- and amphotericin-resistant strains. In particular, this is typical for the species belonging to the genus *Candida*. Candidoses affect about 400 thousand people annually. At the same time, fungi use various biochemical mechanisms to avoid lethal effects of the fungicides; they could use special transport systems to expel these drugs from the cells. Recently, metabolites active against such strains have been found in marine organisms. For example, a polyketide metabolite furosoline A (**30**) was isolated from the bacterium *Actinomadura* sp. using liquid chromatography – mass spectrometry. This bacterium was isolated from the ascidian *Ecteinascidia turbinata*. Furosoline A (**30**) (Fig. 4) inhibits pathogenic fungi at a dose of 16 µg/ml. Successful use of **30** for treatment of mice with experimental multiple candidosis was demonstrated [52].

It is of interest that some marine-derived steroids, such as polar steroids from the sponge *Dysidea arenaria*, increase activity of the antifungal drugs and cancel resistance of pathogenic fungi to fluconazole. At the same time, activity of the antifungal drugs after the treatment with these steroids can be increased 35-fold [53].

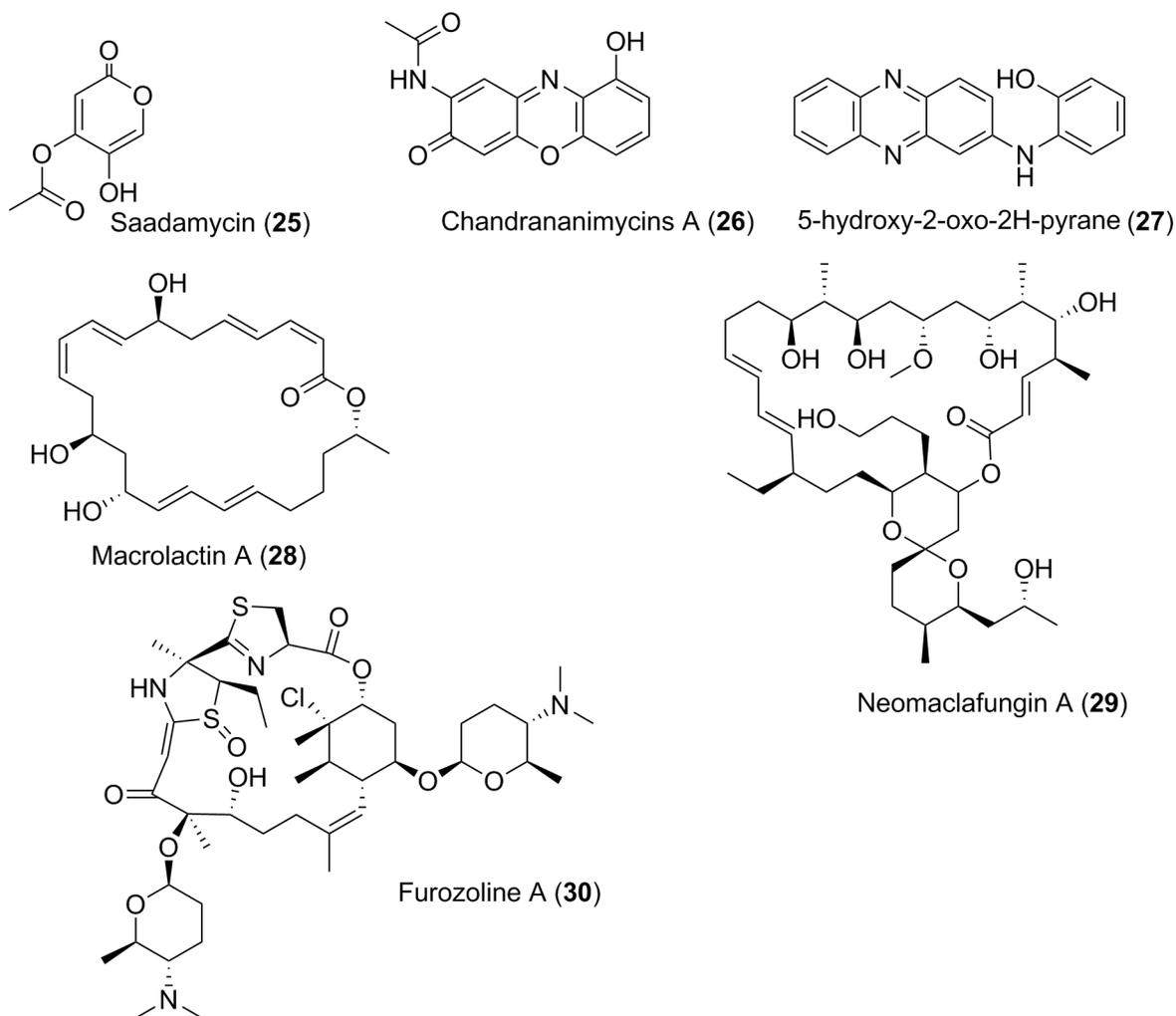


Fig. 3. Some antifungal metabolites from marine microorganisms.

#### ANTIBIOTICS WITH ANTIVIRAL PROPERTIES

Only a few antiviral metabolites were found among the compounds isolated from marine actinobacteria. This may be due to a less intensive search for such substances compared to anti-tumor and anti-fungal natural marine-derived products. Benzastatin C (**31**) (Fig. 4) was obtained from the marine bacterium *Streptomyces nitrosporeus*. It demonstrates a dose-dependent inhibitory effects with  $EC_{50}$  1.92 and 0.53  $\mu\text{g}/\text{ml}$  against HSV-1 and HSV-2 herpes viruses, respectively. Thus, it was more active in these tests than Ara C – a well-known drug that was also created on the basis of marine natural compounds and became the progenitor of the nucleoside-based antiviral drugs. At the same time, cytotoxic properties of **31** were low. Compound **31** shows also antiviral action against the vesicular stomatitis virus. It was shown that the presence of chlorine atom is necessary for the antiviral effect of this leader compound [54].

Antimycin 1A (**32**) (Fig. 4) [55] was isolated from the new species, *Streptomyces kaviengensis*, obtained from the sample of bottom sediment collected near New Ireland (Papua New Guinea). The isolated compound shows an extremely high antiviral effect ( $IC_{50}$  4 nM) against equine encephalitis virus. Its mechanism of action is related to the ability of inhibiting the electron transport network in mitochondria and suppressing *de novo* pyrimidine biosynthesis. Administration of the drug *in vivo* increases survival of the animals infected with this virus. Antimycin inhibits also other RNA viruses.

Adenovirus infections are well known to be associated with high mortality in patients with immunodeficiency. It is commonly recognized that there are no effective medical means to combat such infections. An actinobacterium belonging to the *Streptomyces* genus was isolated from one of fjords off the coast of Norway. It produced simple substances, butenolides, as mixtures of close related metabolites. Butenolide derivatives (**33-37**) (Fig. 4),

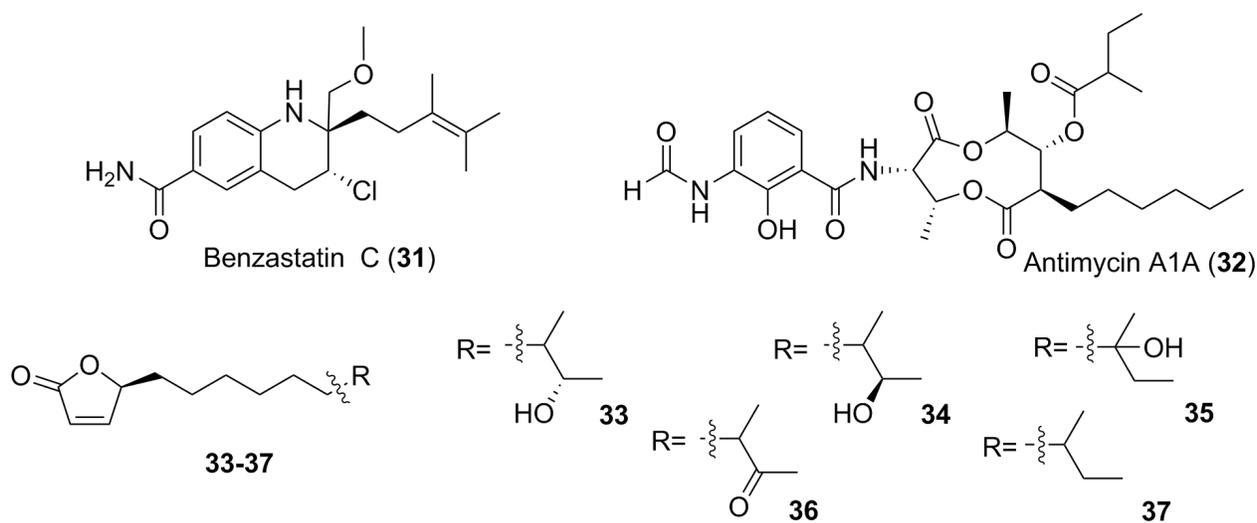


Fig. 4. Some marine antibiotics with antiviral properties.

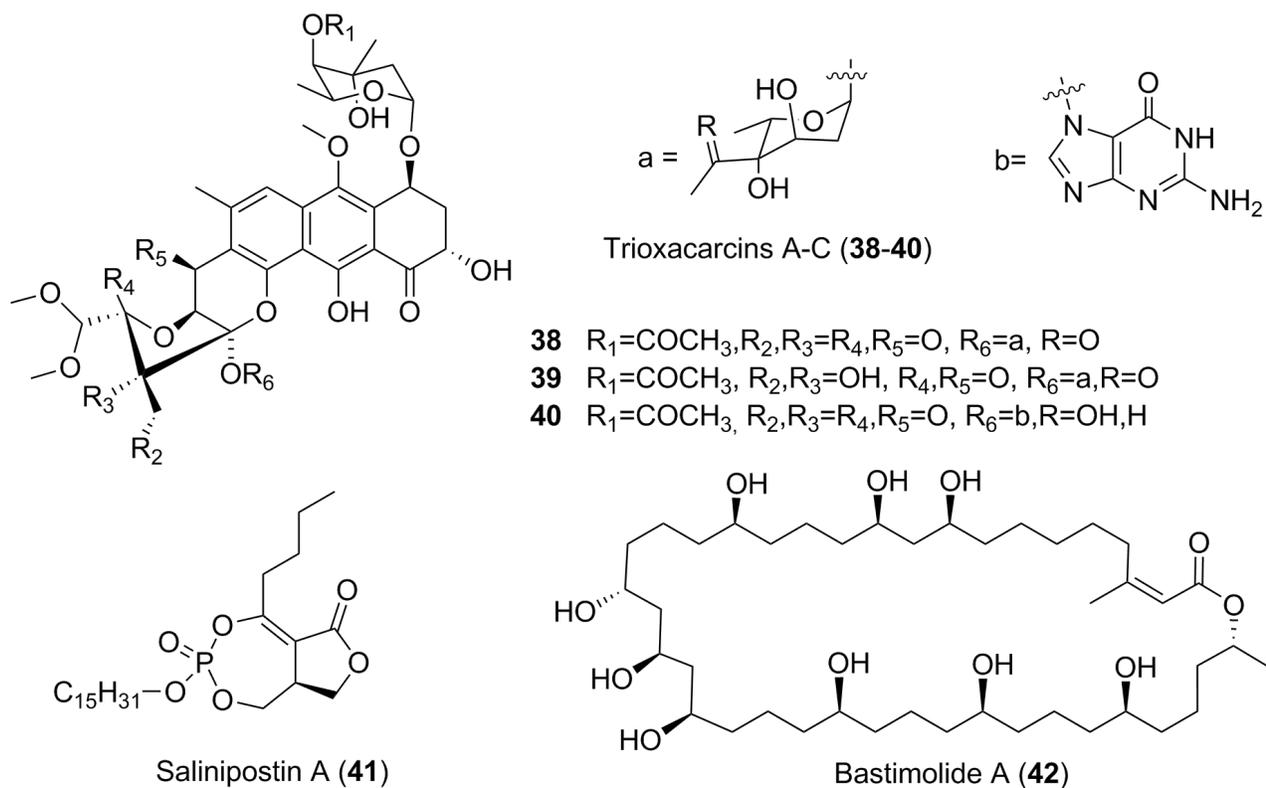


Fig. 5. Some new antimalarial compounds from marine bacteria.

isolated from the extracts of this streptomycete were active against adenoviruses. The most active compound shows an effective  $EC_{50}$  concentration of 91  $\mu M$  and no noticeable cytotoxicity against human cells. These substances are of interest as model compounds for creating appropriate drugs [56].

#### ANTIPARASITIC COMPOUNDS FROM MARINE BACTERIA

For centuries, malaria claimed thousands of lives each year until first and second-generations of antimalarial drugs were developed. Quinine, chloroquine, and

other therapeutics protected many millions of people, but unfortunately, later some forms of malaria resistant to such drugs emerged. As it is well known, this disease is caused by the parasitic protozoan, malarial plasmodium (*Plasmodium falciparum*), which is transferred to people via mosquito bites. In Africa, malaria is the most common disease; increasing annual numbers of malaria cases are detected in South America and South-East Asia. The search for new antimalarial compounds and development of appropriate drugs are becoming more and more important year by year. Recent successes in this scientific field was recognized by awarding the 2015 Nobel prize for creation of a new antimalarial drug artemisinin and new antiparasitic drug ivermectin developed on the basis of the corresponding natural compounds.

Marine-derived antimalarial compounds include, for example, trioxacarcins (**38-40**) (Fig. 5), produced by *Streptomyces ochraceus* and *S. bottropensis*. All the trioxacarcins show high antibacterial, antitumor, and some of them, particularly **38**, antimalarial activities [57]. Salinipostins A-K inhibited the chloroquine-resistant W2 strains of *P. falciparum* with  $EC_{50}$  between 50 nM and 50  $\mu$ M. Of these, salinipostin A shows the strongest effect ( $EC_{50} \sim 50$  nM). It is expected that it would be more difficult to develop resistance to this compound because this drug candidate does not affected heme [58]. Salinipostins were obtained from the microbial strain of *Salinispora* sp. collected from the bottom of the Keawekaheka Bay, Hawaii. Salinipostin A (**41**) inhibits one of the growth stages of plasmodium.

A new polyhydroxymacrolide, bastimolide A (**42**) (Fig. 5), was isolated from the tropical cyanobacterium *Okeania hirsuta*. It inhibited four drug-resistant strains of malaria plasmodium in nanomolar concentrations and was less toxic to both normal and tumor cells [59].

High antimalarial activity was also found for the above-mentioned salinosporamide A (**1**), which has been clinically studied as an antitumor drug. It is assumed that it acts on the 20S proteasome of the parasite. Administration of salinosporamide A at a dose of 130  $\mu$ g/kg protects mice from malaria infection.

## CONCLUSION

Recently, marine-derived natural compounds with antibiotic action have attracted more and more attention. In our country, these compounds are studied by not so many scientific groups, all these groups are concentrated only in two cities, Moscow and Vladivostok. Unexpectedly wide structural diversity of the metabolites from marine bacteria isolates and their promising biological effects makes it quite possible to develop new medical drugs based on them, and several such compounds have already become the active substances of new drugs or are currently undergoing preclinical or clinical trials. Structural nov-

elty and rather complex chemical structure of many of them make it difficult to determine exact structures of such substances, in some cases this task can be solved only with the use of X-ray diffraction analysis. Unlike in the United States and Japan, in Russia targeted total asymmetric synthesis of natural compounds is not developing so well, therefore, additional attention should be given to this scientific direction. Sometimes, it is difficult to obtain the target substances in sufficient quantities for their in-depth study. The problem of obtaining of target substances is solved mainly using one of the two strategies: by creating microbial super producers with selection methods and mutagenesis or by transferring the corresponding gene clusters to easily cultured bacterial species.

Despite the existing difficulties and problems, we have several advantages in Russia when working in this area. First of all, these advantages include: (i) an extended coastline of the country and wide opportunities to collect the necessary biological materials, and (ii) the existence in Vladivostok of the collection of live marine bacteria and fungi, which contains about 4 thousand axenic strains collected in various geographical areas of the World. Nature has created unique structural resources in the marine environment, the study and use of which is an important task of basic research and applied science.

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## REFERENCES

1. URL: <https://www.who.int/ru/news-room/fact-sheets/detail/tuberculosis>.
2. Pendleton, J. N., Gorman, S. P., and Gilmore, B. F. (2013) Clinical relevance of the ESKAPE pathogens, *Exp. Rev. Anti Infect. Ther.*, **11**, 297-308, doi: 10.1586/eri.13.12.
3. URL: <https://www.who.int/ru/news-room/detail/27-02-2017-who-publishes-list-of-bacteria-for-which-new-antibiotics-are-urgently-needed>.
4. Schinke, C., Martins, T., Queiroz, S. C. N., Melo, I. S., and Reyes, F. G. R. (2017) Antibacterial compounds from marine bacteria, 2010-2015, *J. Nat. Prod.*, **80**, 1215-1228, doi: 10.1021/acs.jnatprod.6b00235.
5. Newman, D. J., and Cragg, D. J. (2007) Natural products as sources of new drugs over the last 25 years, *J. Nat. Prod.*, **70**, 461-477, doi: 10.1021/np068054v.
6. Jensen, P. R., and Fenical, W. (1996) Marine bacterial diversity as a resource for novel microbial products, *J. Ind. Microbiology*, **17**, 346-351, doi: 10.1007/BF01574765.
7. Jensen, P. R., and Fenical, W. (1994) Strategies for the discovery of secondary metabolites from marine bacteria: eco-

- logical perspectives, *Annu. Rev. Microbiol.*, **48**, 559-584, doi: 10.1146/annurev.mi.48.100194.003015.
8. Davidson, B. (1995) New dimensions in natural products research: cultured marine microorganisms, *Curr. Opin. Biotechnol.*, **6**, 284-291, doi: 10.1016/0958-1669(95)80049-2.
  9. Pietra, F. (1997) Secondary metabolites from marine microorganisms: bacteria, protozoa, algae and fungi. Achievements and prospects, *Nat. Prod. Rep.*, **14**, 453-464, doi: 10.1039/NP9971400453.
  10. Andryukov, B. G., Mikhailov, V. V., Besednova, N. N., Bynina T. S., and Matosova, E. V. (2018) The bacteriocynogenic potential of marine microorganisms, *Russ. J. Mar. Biol.*, **44**, 433-441, doi: 10.1134/S1063074018060020.
  11. Eom, S. H., Kim, Y. M., and Kim, S. K. (2013) Marine bacteria: potential sources for compounds to overcome antibiotic resistance, *Appl. Microbiol. Biotechnol.*, **97**, 4763-4773, doi: 10.1007/s00253-013-4905-y.
  12. Rahman, H., Austin, B., Mitchell, W. J., Morris, P. S., Jamieson, D. J., et al. (2010) Novel anti-infective compounds from marine bacteria, *Mar. Drugs*, **8**, 498-518, doi: 10.3390/md8030498.
  13. Mikhailov, V. V. (2005) Marine microbiology at PIBOC FEB RAS, *Herald of the Far Eastern Branch Russian Academy of Science*, **2**, 145-152.
  14. Romanenko, L. A., Uchino, M., Frolova, G. M., Tanaka, N., Kalinovskaya, N. I., Latyshev, N., and Mikhailov, V. V. (2007) *Sphingomonas molluscorum* sp. nov., a novel marine isolate with antimicrobial activity, *Int. J. Syst. Evol. Microbiol.*, **57**, 358-363, doi: 10.1099/ijs.0.64441-0.
  15. Manivasagan, P., Venkatesan, S., Sivakumar, S., and Kim, S. K. (2014) Pharmaceutically active secondary metabolites of marine actinobacteria, *Microbiol. Res.*, **169**, 262-278, doi: 10.1016/j.micres.2013.07.014.
  16. Ul Hassan, S. S., Anjum, K., Abbas, S. Q., Akhter, N., Shagufta, B. I., Shah, S. A. A., and Tasneem, U. (2017) Emerging biopharmaceuticals from marine actinobacteria, *Environ. Toxicol. Pharmacol.*, **49**, 34-47, doi: 10.1016/j.etap.2016.11.015.
  17. Weyland, H. (1969) Actinomycetes in North Sea and Atlantic Ocean sediments, *Nature*, **223**, 858, doi: 10.1038/223858a0.
  18. Thomas, T. R. A., Kavlekar, D. P., and LokaBharathi, P. A. (2010) Marine drugs from sponge-microbe association – a review, *Mar. Drugs*, **8**, 1417-1468, doi: 10.3390/md8041417.
  19. Reddy, T. B. K., Thomas, A. D., Stamatis, D., Bertsch, J., Isbandi, M., et al. (2014) The genome online database (GOLD) v.5: a metadata management system based on a four level (meta)genome project classification, *Nucleic Acids Res.*, **43**, 1099-1106, doi: 10.1093/nar/gku950.
  20. Khan, S. T., Tamura, T., Takagi, M., and Shin-ya, K. (2010) *Streptomyces tateyamensis* sp. nov., *Streptomyces marinus* sp. nov. and *Streptomyces haliclona* sp. nov., isolated from the marine sponge *Haliclona* sp., *Int. J. Syst. Evol. Microbiol.*, **60**, 2775-2779, doi: 10.1099/ijs.0.019869-0.
  21. Tian, X. P., Tang, S. K., Dong, J. D., Zhang, Y. Q., Xu, L. H., Zhang, S., and Li, W. J. (2009) *Marinactinospora thermotolerans* gen. nov., sp. nov., a marine actinomycete isolated from a sediment in the northern South China Sea, *Int. J. Syst. Evol. Microbiol.*, **59**, 948-952, doi: 10.1099/ijs.0.005231-0.
  22. Schumann, P., Cui, X., Stackebrandt, E., Kroppenstedt, R. M., Xu, L., and Jiang, C. (2004) *Jonesia quinghaiensis* sp. nov., a new member of the suborder Micrococccineae, *Int. J. Syst. Evol. Microbiol.*, **54**, 2181-2184, doi: 10.1099/ijs.0.63223-0.
  23. Han, S. K., Nedashkovskaya, O. I., Mikhailov, V. V., Kim, S. B., and Bae, K. S. (2003) *Salinibacterium amurskyense* gen. nov., sp. nov., a novel genus of the family Microbacteriaceae from the marine environment, *Int. J. Syst. Evol. Microbiol.*, **53**, 2061-2066, doi: 10.1099/ijs.0.02627-0.
  24. Tian, X. P., Zhi, X. Y., Qiu, Y. Q., Zhang, Y. Q., Tang, S. K., Xu, L. H., Zhang, S., and Li, W. J. (2009) *Sciscionella marina* gen. nov., sp. nov., a marine actinomycete isolated from a sediment in the northern South China Sea, *Int. J. Syst. Evol. Microbiol.*, **59**, 222-228, doi: 10.1099/ijs.0.001982-0.
  25. Maldonado, L. A., Fenical, W., Jensen, P. R., Kauffman, C. A., Mincer, T. J., Ward, A. C., Bull, A. T., and Goodfellow, M. (2005) *Salinispora arenicola* gen. nov., sp. nov. and *Salinispora tropica* sp. nov. obligate marine actinomycetes belonging to the family Micromonosporaceae, *Int. J. Syst. Evol. Microbiol.*, **55**, 1759-1766, doi: 10.1099/ijs.0.63625-0.
  26. Jensen, P. R., Moore, B. S., and Fenical, W. (2015) The marine actinomycete genus *Salinispora*: a model organism for secondary metabolite discovery, *Nat. Prod. Rep.*, **32**, 738-751, doi: 10.1039/c4np00167b.
  27. Feling, R. H., Buchanan, G. O., Mincer, T. J., Kauffman, C. A., Jensen, P. R., and Fenical, W. (2003) Salinosporamide A: A highly cytotoxic proteasome inhibitor from a novel microbial source, a marine bacterium of the new genus *Salinispora*, *Angew. Chem. Int. Ed.*, **42**, 355-357, doi: 10.1002/anie.200390115.
  28. Williams, P. G., Buchanan, G. O., Feling, R. H., Kauffman, C. A., Jensen, P. R., and Fenical, W. (2005) New cytotoxic salinosporamides from the marine actinomycete *Salinispora tropica*, *J. Org. Chem.*, **70**, 6196-6203, doi: 10.1021/jo050511+.
  29. Macherla, V. R., Mitchell, S. S., Manam, R. R., Reed, K. A., Chao, T. H., et al. (2005) Structure-activity relationship studies of salinosporamide A (NPI-0052), a novel marine derived proteasome inhibitor, *J. Med. Chem.*, **48**, 3684-3687, doi: 10.1021/jm048995+.
  30. Miller, E. D., Kauffman, C. A., Jensen, P. R., and Fenical, W. (2007) Piperazimycins: cytotoxic hexadepsipeptides from a marine-derived bacterium of the genus *Streptomyces*, *J. Org. Chem.*, **72**, 323-330, doi: 10.1021/jo061064g.
  31. Kalinovskaya, N. I., Romanenko, L. A., Kalinovskaya, A. I., Ermakova, S. P., Dmitrenok, P. S., and Afiyatullo, S. S. (2017) The antitumor antibiotics complex of aureolic acids from the marine sediment-associated strain of *Streptomyces* sp. KMM 9048, *Nat. Prod. Comm.*, **12**, 571-577.
  32. Rinehart, K. L. (1999) Antitumor compounds from tunicates, *Med. Res. Revs.*, **20**, 1-27, doi: 10.1002/(SICI)1098-1128(200001)20:1<1::AID-MED1>3.0.CO;2-A.
  33. Rath, C. M., Janto, B., Earl, J., Ahmed, A., Hu, F. Z., et al. (2011) Meta-omic characterization of the marine invertebrate microbial consortium that produces the chemotherapeutic natural product ET-743, *ACS Chem. Biol.*, **6**, 1244-1256, doi: 10.1021/cb200244t.
  34. Blockley, A., Elliott, D. R., Roberts, A. P., and Sweet, M. (2017) Symbiotic microbes from marine invertebrates: driving a new era of natural product drug discovery, *Diversity*, **9**, 49, doi: 10.3390/d9040049.

35. Bibi, F., Faheem, M., Azhar, E. I., Yasir, M., Alvi, S. A., Kamal, M. A., Ullah, I., and Naseer, M. I. (2017) Bacteria from marine sponges: a source of new drugs, *Curr. Drug Metab.*, **18**, 11-18, doi: 10.2174/1389200217666161013090610.
36. Francisco, J. A., Cervený, C. G., Meyer, D. L., Mixan, B. J., Klussman, K., et al. (2003) cAC10-vcMMAE, an anti-CD30–monomethyl auristatin E conjugate with potent and selective antitumor activity, *Blood*, **102**, 1458-1465, doi: 10.1182/blood-2003-01-0039.
37. Zhou, Q., and Kim, J. (2015) Advances in the development of site-specific antibody-drug conjugation, *Anticancer Agents Med. Chem.*, **15**, 828-836, doi: 10.2174/1871520615666150302125448.
38. Jang, K. H., Nam, S. J., Locke, J. B., Kauffman, C. A., Beatty, D. S., Paul, L. A., and Fenical, W. (2013) Anthracimycin, a potent anthrax antibiotic from a marine-derived actinomycete, *Angew. Chem. Int. Ed.*, **52**, 7822-7824, doi: 10.1002/anie.201302749.
39. Alt, S., and Wilkinson, B. (2015) Biosynthesis of the novel macrolide antibiotic anthracimycin, *ACS Chem. Biol.*, **10**, 2468-2479, doi: 10.1021/acschembio.5b00525.
40. Socha, A. M., LaPlante, K. L., and Rowley, D. C. (2006) New bisanthraquinone antibiotics and semi-synthetic derivatives with potent activity against clinical *Staphylococcus aureus* and *Enterococcus faecium* isolates, *Bioorg. Med. Chem.*, **14**, 8446-8454, doi: 10.1016/j.bmc.2006.08.038.
41. McArthur, K. A., Mitchell, S. S., Tsueng, G., Rheingold, A., White, D. J., Grodberg, J., Lam, K. S., and Potts, B. C. M. (2008) Lynamycins A-E, chlorinated bisindole pyrrole antibiotics from a novel marine actinomycete, *J. Nat. Prod.*, **71**, 1732-1737, doi: 10.1021/np800286d.
42. Tareq, F. S., Kim, J. H., Lee, M. A., Lee, H. S., Lee, J. S., Lee, Y. J., and Shin, H. J. (2013) Antimicrobial gageo-macrolactins characterized from the fermentation of the marine-derived bacterium *Bacillus subtilis* under optimum growth conditions, *J. Agricult. Food Chem.*, **61**, 3428-3434, doi: 10.1021/jf4009229.
43. Palomo, S., González, I., de la Cruz, M., Martín, J., Tormo, J. R., et al. (2013) Sponge-derived *Kocuria* and *Micrococcus* spp. as sources of the new thiazolyl peptide antibiotic kocurin, *Mar. Drugs*, **11**, 1071-1086, doi: 10.3390/md11041071.
44. Mahajan, G., Thomas, B., Parab, R., Patel, Z. E., Kuldharan, S., et al. (2013) *In vitro* and *in vivo* activities of antibiotic PM181104, *Antimicrob. Agents Chemother.*, **57**, 5315-5319, doi: 10.1128/AAC.01059-13.
45. El-Gendy, M. M. A., and El-Bondkly, A. M. A. (2010) Production and genetic improvement of a novel antimycotic agent, saadamycin, against dermatophytes and other clinical fungi from endophytic *Streptomyces* sp. Hedaya48, *J. Indust. Microbiol. Biotech.*, **37**, 831-841, doi: 10.1007/s10295-010-0729-2.
46. Maskey, R. P., Li, F. C., Qin, S., Fiebig, H. H., and Laatsch, H. (2003) Chandranamycins A approximately C: production of novel anticancer antibiotics from a marine *Actinomadura* sp. isolate M048 by variation of medium composition and growth conditions, *J. Antibiot.*, **56**, 622-629, doi: 10.7164/antibiotics.56.622.
47. Gao, X., Lu, Y., Xing, Y., Ma, Y., Lu, J., Bao, W., Wang, Y., and Xi, T. (2012) A novel anticancer and antifungus phenazine derivative from a marine actinomycete BM-17, *Microbial. Res.*, **167**, 616-622, doi: 10.1016/j.micres.2012.02.008.
48. Nagao, T., Adachi, K., Sakai, M., Nishima, M., and Sano, H. (2001) Novel macrolactins as antibiotic lactones from a marine bacterium, *J. Antibiot.*, **54**, 333-339, doi: 10.7164/antibiotics.54.333.
49. Karpinsky, T. M. (2019) Marine macrolides with antibacterial and/or antifungal activity, *Mar. Drugs*, **17**, 241, doi: 10.3390/md17040241.
50. Sato, S., Iwata, F., Yamada, S., and Katayama, M. (2012) Neomaclafungins A–I: oligomycin-class macrolides from a marine-derived actinomycete, *J. Nat. Prod.*, **75**, 1974-1982, doi: 10.1021/np300719g.
51. Zhu, S., and Wu, Y. (2017) Synthesis and configuration of neomaclafungin A, *Chem. Asian J.*, **12**, 2211-2215, doi: 10.1002/asia.201700950.
52. Wyche, T. P., Piotrowsky, J. S., Hou, Y., Braun, D., Deshpande, R., et al. (2014) Furozoline A: marine-derived polyketide with antifungal *in vivo* efficacy, *Angew. Chem. Int. Ed.*, **53**, 1-5, doi: 10.1002/anie.201405990.
53. Abdelmohsen, U. R., Balasubramanian, S., Oelschlaeger, T. A., Grkovic, T., Pham, N. B., Quinn, R. J., and Hentschel, U. (2017) Potential of marine natural products against drug-resistant, viral, and parasitic infections, *Lancet Infect. Dis.*, **17**, e30-e41, doi: 10.1016/S1473-3099(16)30323-1.
54. Lee, J. G., Yoo, I. D., and Kim, W. G. (2007) Differential antiviral activity of benzastatin C and its dechlorinated derivative from *Streptomyces nitrosporeus*, *Biol. Pharm. Bull.*, **30**, 795-797, doi: 10.1248/bpb.30.795.
55. Raveh, A., Delecta, P. C., Dobry, C. J., Peng, W., Schultz, P. J., Blakely, P. K., and Miller, D. J. (2013) Discovery of potent broad spectrum antivirals derived from marine actinobacteria, *PLoS One*, **8**, e 82318, doi: 10.1371/journal.pone.0082318.
56. Strand, M., Carlsson, M., Uvell, H., Islam, K., Edlung, K., Cullman, I., et al. (2014) Isolation and characterization of anti-adenoviral secondary metabolites from marine actinobacteria, *Mar. Drugs*, **12**, 799-821, doi: 10.3390/md12020799.
57. Maskey, R. P., Helmke, E., Kayser, O., Fiebig, H. H., Maier, A., Busche, A., and Laatsch, H. (2004) Anti-cancer and antibacterial trioxacarcins with high anti-malaria activity from a marine *Streptomyces* and their absolute stereochemistry, *J. Antibiot.*, **57**, 771-779, doi: 10.7164/antibiotics.57.771.
58. Schulze, C. J., Navarro, G., Ebert, D., DeRisi, J., and Lenington, R. G. (2015) Salinipostins A-K, long-chain bicyclic phosphotriesters as a potent and selective anti-malarial chemotype, *J. Org. Chem.*, **80**, 1312-1320, doi: 10.1021/jo5024409.
59. Shao, C. L., Lenington, R. G., Balunas, M. J., Centeno, A., Boudreau, P., et al. (2015) Bastimolide A, a potent anti-malarial polyhydroxy macrolide from a marine cyanobacterium *Okeania hirsuta*, *J. Org. Chem.*, **80**, 7849-7855, doi: 10.1021/acs.joc.5b01264.